

November 14, 2024

John Murray
Ashland Public Schools
87 West Union St.
Ashland, MA 01721

Re: Limited Mold Assessment & Sampling
Mindess Elementary School, 90 Concord St., Ashland, MA
FLI Project # 24-2539

Dear Mr. Murray:

On November 6, 2024, FLI Environmental, Inc. (FLI) conducted a limited mold assessment and air sampling at the above-mentioned address. The purpose of this survey was to address potential indoor air quality concerns regarding mold. The following letter summary represents the assessment including our scope of work, sampling methodology, discussion of results and conclusion.

I. SCOPE OF WORK

1. Collection of air sample(s) within the subject areas for airborne mold;
2. Preparation of a summary report detailing the sampling methodology along with analytical results, a discussion of the results, and a conclusion.

II. SAMPLING METHODOLOGY

1. Airborne Sampling for Mold:

The concentration and identification of the genera of airborne mold was performed through the use of Air-O-Cell cassettes. This method utilizes an air pump to draw air at a predetermined flow rate through a spore trap cassette containing a slide coated with an optically-transparent adhesive. Airborne particulate, including spores is impacted onto the slide, and then submitted to the laboratory where it is stained and analyzed by optical microscopy at magnifications between 200X and 1000X. Air-O-Cell samples collected at the above referenced location were enumerated and speciated by SanAir Technologies Laboratory located in North Chesterfield, VA.

This method does not differentiate between viable and non-viable fungal spores. In addition, this technique does not allow for the differentiation between *Aspergillus* and *Penicillium* spores. Other non-distinctive spores are reported in categories such as *Ascospores* or *Basidiospores*.

III. DISCUSSION OF RESULTS

1. Airborne Sampling for Mold:

Although there are currently no standards or regulations to indicate acceptable levels of airborne fungal spores derived from indoor environments, a comparison of the indoor/outdoor (I/O) ratio of total spore enumeration is recommended below 1.0 (indoor levels should not overly exceed outdoor levels). The indoor and outdoor spore types and distribution should also be similar. According to ACGIH, “... differences that can be detected with manageable sample sizes are likely to be in 10-fold multiplicative steps (e.g., 100 versus 1000...)”. Following this logic, if total fungal spores are ten (10) times greater in the sample from a suspect area than in the negative control sample collected from a non-suspect area, then that sample area may be a fungal amplification site.

Five (5) samples were collected on November 6, 2024, inside the building. An ambient sample was collected outside for comparison with the indoor sample. The dominant genus in the indoor samples with higher mold concentrations was observed as *Smuts/Myxomycetes*. They share similar microscopic morphology and are typically grouped together because they are both associated with plants and the outdoors. Smuts are ubiquitous and associated with cereal crops, grasses, weeds and other flowering plants. Myxomycetes are common in forests, often developing on tree bark, decaying logs, stumps, dead leaves, and other organic material. Similar to true molds, they are mostly associated with cool and moist habitats. Smuts can cause allergens, including type I allergies like hay fever and asthma. Myxomycetes are not known to be pathogenic and associations between Myxomycetes and humans are rare, usually nothing to worry about. *See Appendix for Air-O-Cell Sample Results.*

IV. CONCLUSIONS & RECOMMENDATIONS

Indoor air quality problems are often the result of complex and dynamic interactions between building systems, space use activities, management practices and occupant expectations. In most cases, indoor contaminants become problematic through irritation or odor before they reach levels toxic to humans. An HVAC system that brings fresh air into the space will generally remove pollutants from the occupied space and dilute the levels of pollutants in that space. With reduced exposure to pollutants, a reduction or elimination of symptoms in occupants should occur.

When an indoor air investigation is conducted, the goal is to determine whether the detected contaminant concentrations originate from the occupied spaces being surveyed or if they merely represent typical background concentrations. The dominant species identified (*Smuts/Myxomycetes*) are associated with plants and not typically found indoors, however classrooms with plants or flowers are likely contributors to elevated airborne Smuts/Myxomycetes levels. Decaying leaves typical to the season are also likely contributors to elevated levels.

1. *Based on analytical data of samples collected, the levels of Smuts/Myxomycetes found in the samples in the subject areas were elevated relative to the ambient*

sample, but do not suggest a fungal amplification concern. However, sensitized individuals may experience symptoms associated with allergies.

2. *Elevated levels are likely attributable to indoor plants in the subject areas and/or seasonal increases associated with decaying leaves contributing to elevated levels in ambient air entrained into the subject areas. If occupants are experiencing symptoms, removal of indoor plants and cleaning of affected areas using a biocide is recommended, with follow-up air sampling upon completion.*

V. LIMITATIONS AND CONDITIONS OF THIS REPORT

The recommendations and conclusions discussed herein are based solely and in reliance upon information collected as a result of the activities delineated in the Proposal. FLI neither attests nor renders an opinion as to the accuracy or comprehensiveness of the analytical results. There is a limit to all investigations of this type in the sense that the researcher must draw conclusions and develop recommendations with information obtained from research, site evaluation and limited sampling and analysis. FLI does not render any warranty, either express or implied, as to the conditions of the Site beyond that observed during the Site survey. The passage of time may also result in a change in the characteristics at the Site. FLI does not render an opinion as to conditions which may change subsequent to the date of the Site reconnaissance. FLI does not render an opinion as to conditions at uninspected or obstructed portions of the Site (e.g. ceiling plenums or air handling equipment), or those areas not sampled as part of this survey. FLI performed professional services and rendered conclusions in accordance with generally accepted practices of other environmental consultants undertaking similar investigations at the same time in the same geographical area. FLI exercised the degree of care and skill generally exercised by other environmental consultants under similar circumstances and conditions.

Thank you for choosing FLI to assist you on this project. I hope the information that we provide in this report fulfills your requirements. If you have any questions about the information contained herein, please do not hesitate to contact me at (781) 251-0040.

Sincerely,
FLI Environmental, Inc.



Samantha Claus
Project Manager

APPENDIX

AIR SAMPLE ANALYTICAL RESULTS



Name: FLI Environmental, Inc.
Address: 69 Bridge Street
 Dedham, MA 02026
Phone: 781-251-0040

Project Number: 24-2539
P.O. Number:
Project Name: 90 Concord St. Ashland, MA
Collected Date: 11/6/2024
Received Date: 11/7/2024 10:50:00 AM

SanAir ID Number
24064184
 FINAL REPORT
 11/11/2024 5:44:47 PM

Analyst: Willis, Madeline

Air Cassette Analysis - Spores % of Outside Air



593%
A

Count/m ³ higher than Baseline	Smuts/Myxomycetes
Count/m ³ comparable to Baseline	
Within 50% of Baseline Count/m ³	

*The Baseline Level (100%) represents the average baseline sample counts. Counts above the baseline may indicate higher than expected levels of a given result.



Name: FLI Environmental, Inc.
Address: 69 Bridge Street
 Dedham, MA 02026
Phone: 781-251-0040

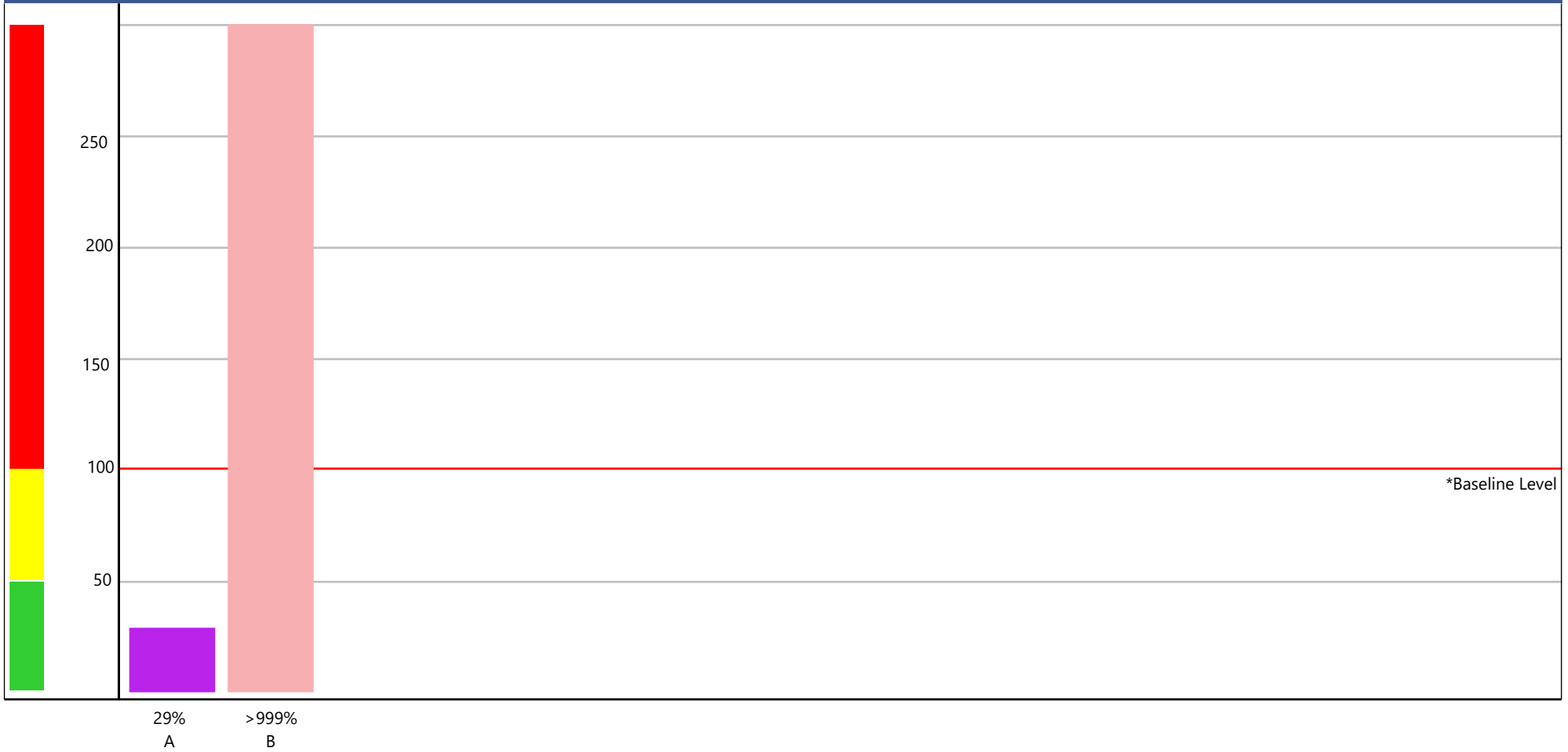
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Analyst: Willis, Madeline

Air Cassette Analysis - Spores % of Outside Air

SanAir ID : 24064184-2 Sample # : 2 ID : Classroom Rm 167



■ Count/m ³ higher than Baseline	A Cladosporium species	B Smuts/Myxomycetes
■ Count/m ³ comparable to Baseline		
■ Within 50% of Baseline Count/m ³		

*The Baseline Level (100%) represents the average baseline sample counts. Counts above the baseline may indicate higher than expected levels of a given result.



Name: FLI Environmental, Inc.
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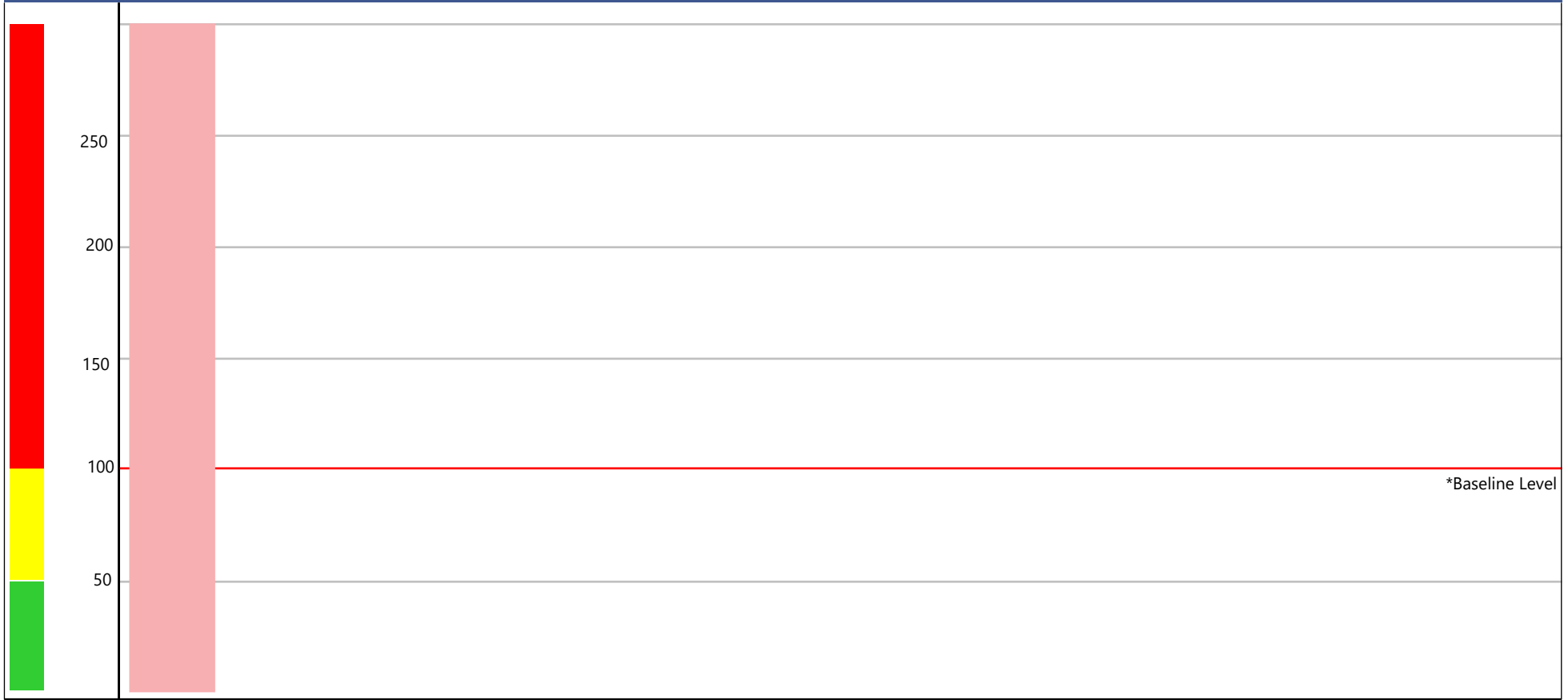
Project Number: 24-2539
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Analyst: Willis, Madeline

Air Cassette Analysis - Spores % of Outside Air

SanAir ID : 24064184-3 Sample # : 3 ID : Classroom Rm 145



544%
A

Count/m³ higher than Baseline	Smuts/Myxomycetes
Count/m³ comparable to Baseline	
Within 50% of Baseline Count/m³	

*The Baseline Level (100%) represents the average baseline sample counts. Counts above the baseline may indicate higher than expected levels of a given result.



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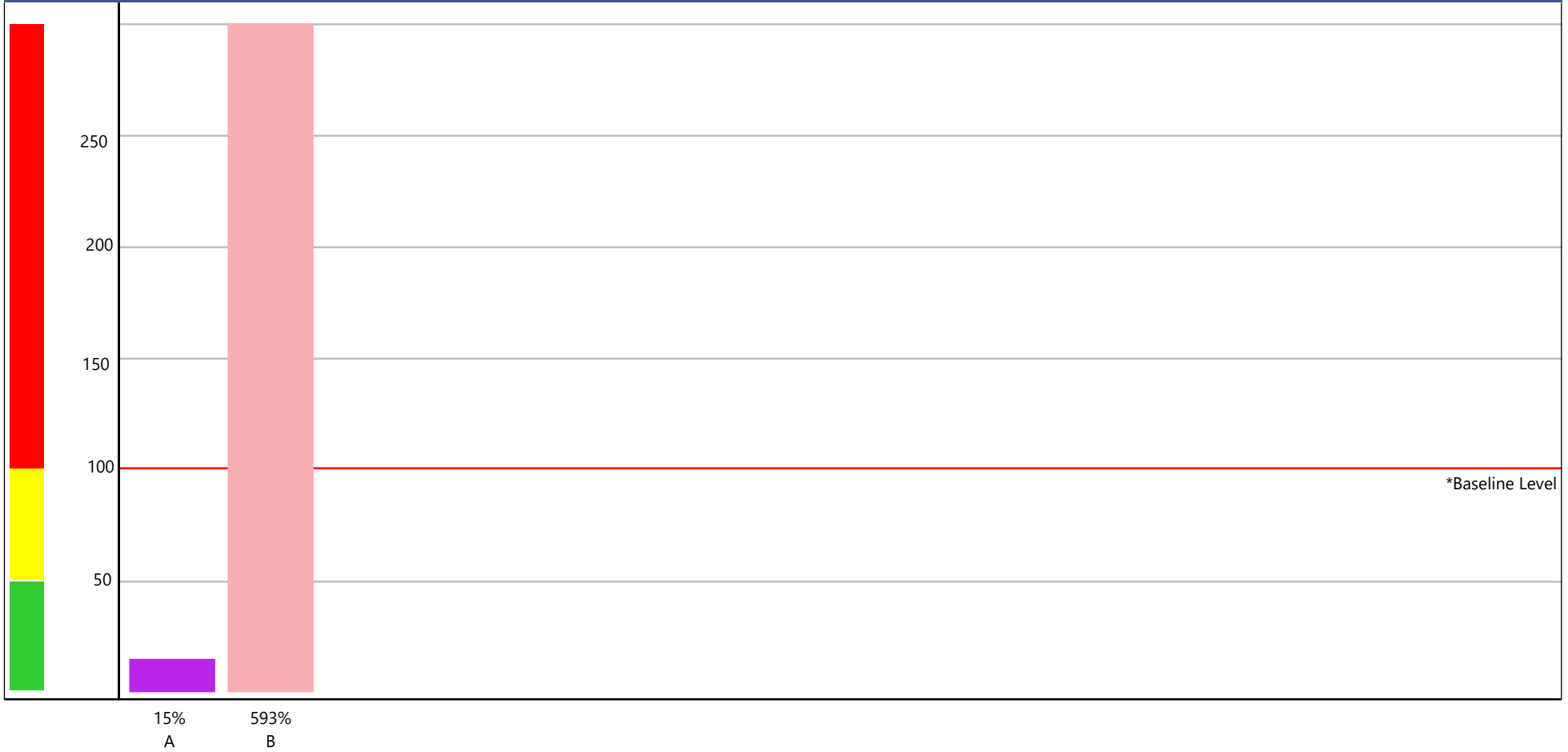
Project Number: 24-2539
P.O. Number:
Project Name: 90 Concord St. Ashland, MA
Collected Date: 11/6/2024
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SanAir ID Number
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Analyst: Willis, Madeline

Air Cassette Analysis - Spores % of Outside Air

SanAir ID : 24064184-4 Sample # : 4 ID : Classroom Rm 154



Count/m ³ higher than Baseline	Cladosporium species	Smuts/Myxomycetes
Count/m ³ comparable to Baseline		
Within 50% of Baseline Count/m ³		

*The Baseline Level (100%) represents the average baseline sample counts. Counts above the baseline may indicate higher than expected levels of a given result.



Name: FLI Environmental, Inc.
Address: 69 Bridge Street
 Dedham, MA 02026
Phone: 781-251-0040

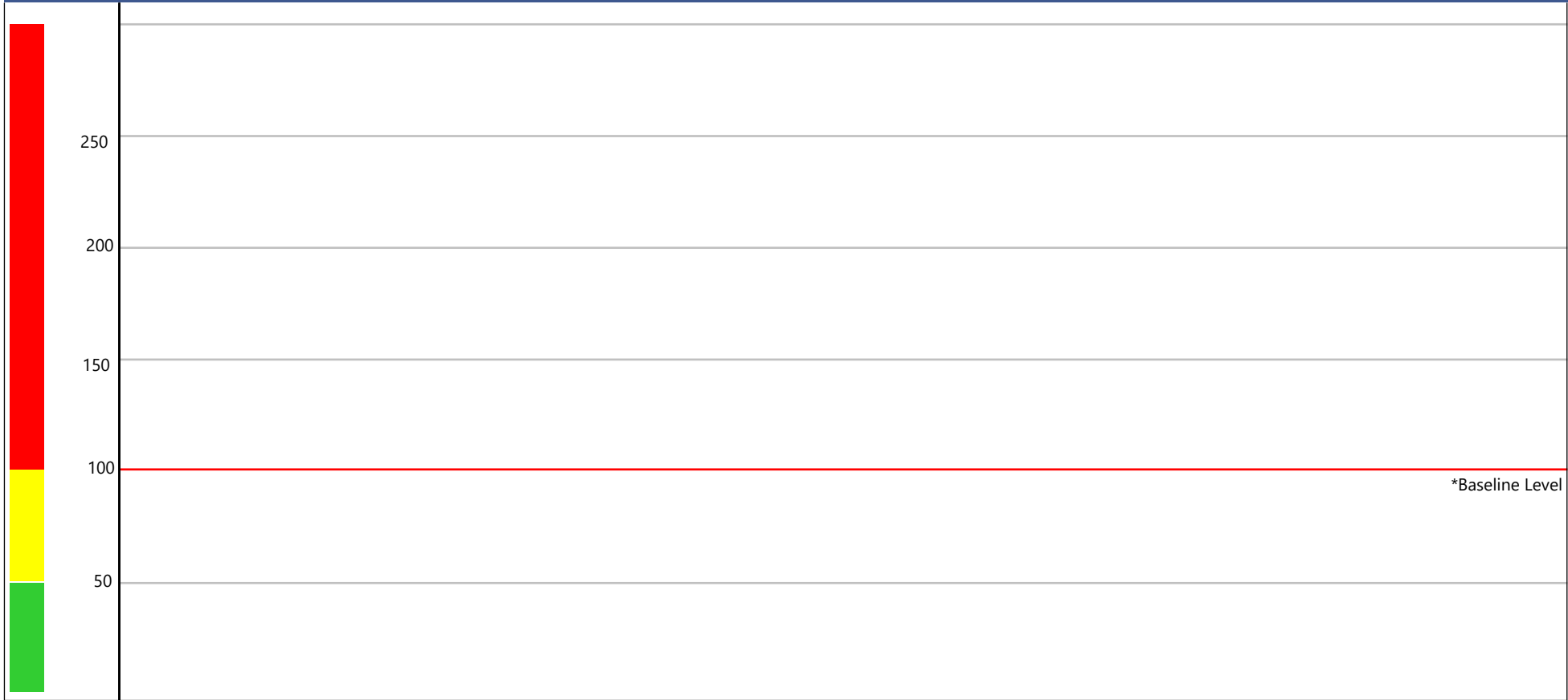
Project Number: 24-2539
P.O. Number:
Project Name: 90 Concord St. Ashland, MA
Collected Date: 11/6/2024
Received Date: 11/7/2024 10:50:00 AM

SanAir ID Number
24064184
 FINAL REPORT
 11/11/2024 5:44:47 PM

Analyst: Willis, Madeline

Air Cassette Analysis - Spores % of Outside Air

SanAir ID : 24064184-5 Sample # : 5 ID : Classroom Rm 149



- Count/m³ higher than Baseline
- Count/m³ comparable to Baseline
- Within 50% of Baseline Count/m³

No organisms to graph. Normalized organism counts may not have exceeded the organism thresholds, or there were no organism counts for this sample. Please refer to the analysis report.

*The Baseline Level (100%) represents the average baseline sample counts. Counts above the baseline may indicate higher than expected levels of a given result.



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Project Number: 24-2539
P.O. Number:
Project Name: 90 Concord St. Ashland, MA
Collected Date: 11/6/2024
Received Date: 11/7/2024 10:50:00 AM

Organism Descriptions

The descriptions of the organisms presented are derived from various reference materials. The laboratory report is based on the data derived from the samples submitted and no interpretation of the data, as to potential, or actual, health effects resulting from exposure to the numbers of organisms found, can be made by laboratory personnel. Any interpretation of the potential health effects of the presence of this organism must be made by qualified professional personnel with first hand knowledge of the sample site, and the problems associated with that site.

Mycelial Fragments - A mycelium (plural = mycelia) is the "body" of a fungus. It is a collective term for hyphae (singular = hypha), which are the tubular units of the mycelium usually composed of chitin. The terms hyphae and mycelial fragments are used interchangeably. [This information was referenced from the mycology text "The Fifth Kingdom"] In some cases a fungal identification cannot be obtained due to lack of sporulation. Only the mycelial fragments are present, and cannot be identified without the distinguishing characteristics of the spores or the structures they grow from.

Health Effects: Allergic reactions may occur in the presence of spores (conidia) or mycelial/hyphal fragments.

Alternaria species - This genus comprises a large number of saprobes and plant pathogens. It is one of the predominate airborne fungal spores indoor and outdoor. Outdoors it may be isolated from samples of soil, seeds, and plants. It is one of the more common fungi found in nature, extremely widespread and ubiquitous. Conidia are easily carried by the wind, with peak concentrations in the summer and early fall. It is commonly found in outdoor samples. It is often found in indoor environments, on drywall, ceiling tiles, in house dust, carpets, textiles, and on horizontal surfaces in building interiors. Often found on window frames. This genus also includes species that were once identified as *Ulocladium*. Genetic testing has shown that the two are not phylogenetically distinct, and as such have been combined.

Health Effects: In humans, it is recognized to cause allergic responses. Because of the large size of the spores, it can be deposited in the nose, mouth and upper respiratory tract, causing nasal septum infections. It has been known to cause Baker's asthma, farmer's lung, and hay fever. It has been associated with hypersensitivity pneumonitis, sinusitis, dermatomycosis, onychomycosis, subcutaneous phaeohyphomycosis, and invasive infection. Common cause of extrinsic asthma (immediate-type hypersensitivity: type I). Acute symptoms include edema and bronchospasms, chronic cases may develop pulmonary emphysema.

References: Flannigan, Brian, Robert A. Samson, and J. David Miller, eds. *Microorganisms in Home and Indoor Work Environments*. London and NY: Taylor & Francis, 2001. de Hoog, G.S. et al. *Atlas of Clinical Fungi*. 4th ed. Foundation Atlas of Clinical Fungi. 2020

Ascospores - From the fungal Subphylum Ascomycotina. Ascospores are ubiquitous in nature and are commonly found in the outdoor environment. This class contains the "sac fungi" and yeasts. Some ascospores can be identified by spore morphology, however; some care should be exercised with regard to specific identification. They are identified on tape lifts and non-viable analysis by the fact that they have no attachment scars and are sometimes enclosed in sheaths with or without sacs.

Ascomycetes may develop both sexual and asexual stages. Rain and high humidity may help asci to release, and disperse ascospores, which is why during these weather conditions there is a great increase in counts.

Health Effects: This group contains possible allergens.

Aspergillus/Penicillium - These spores are easily aerosolized. Only through the visualization of reproductive structures can the genera be distinguished. Also included in this group may be spores of the genera *Acremonium*, *Phialophora*, *Verticillium*, *Paecilomyces*, *Talaromyces* etc. Small, round to ovoid spores of this group lack the necessary distinguishing characteristics when seen on non-viable examination.

Health Effects: Can cause a variety of symptoms including allergic reactions. Most symptoms occur if the individual is immunocompromised in some way (HIV, cancer, etc).

Basidiospores - From the Subphylum Basidiomycotina which contains the mushrooms, shelf fungi, and a variety of other macrofungi. They are saprophytes, ectomycorrhizal fungi or agents of wood rot, which may destroy the structure wood of buildings. It is extremely difficult to identify a specific genera of mushrooms by using standard culture plate techniques. Some basidiomycete spores can be identified by spore morphology; however, some care should be exercised with regard to specific identification. The release of basidiospores is dependant upon moisture, and they are dispersed by wind.

Health Effects: Many have the potential to produce a variety of toxins. Members of this group may trigger Type I and III fungal hypersensitivity reactions. Rarely reported as opportunistic pathogens.



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Cladosporium species - The most commonly identified outdoor fungus. The outdoor numbers are reduced in the winter and are often high in the summer. Often found indoors in numbers less than outdoor numbers. It is commonly found on the surface of fiberglass duct liner in the interior of supply ducts. A wide variety of plants are food sources for this fungus. It is found on dead plants, woody plants, food, straw, soil, paint and textiles. Often found in dirty refrigerators and especially in reservoirs where condensation is collected, on moist window frames it can easily be seen covering the whole painted area with a velvety olive green layer.

Health Effects: It is a common allergen. It can cause mycosis. Common cause of extrinsic asthma. Acute symptoms include edema and bronchospasms, chronic cases may develop pulmonary emphysema. Illnesses caused by this genus can include phaeohyphomycosis, chromoblastomycosis, hay fever and common allergies.

References: Flannigan, Brian, Robert A. Samson, and J. David Miller, eds. Microorganisms in Home and Indoor Work Environments. London and NY: Taylor & Francis, 2001. de Hoog, G.S. et al. Atlas of Clinical Fungi. 4th ed. Foundation Atlas of Clinical Fungi. 2020

Rusts - From the group Uredinales, called Rusts due to the color of the spores, which are known for causing disease in plants.

Smuts/Myxomycetes - Smuts and Myxomycetes are parasitic plant pathogens. They are typically grouped together due to their association with plants, the outdoors and because they share similar microscopic morphology.

Health Effects: Can produce type I fungal hypersensitivity reactions.

References: Martin, G.W., C.J. Alexopoulos, and M.L. Farr. The Genera of Myxomycetes. Iowa City, Iowa: University of Iowa Press, 1983.

Additional Information

Air Cassette Analyses

Air cassette reports indicate the genus and concentration of viable (living) and non-viable mold spores detected on the slide (A2 Analysis). Whether or not these spores are viable cannot be determined using this type of analysis. However, keep in mind that spores can remain allergenic even after cellular death. Other possible allergens include dander, pollen and fibers which are included in air cassette reports for the A1 Analysis. A1 and A2 analyses are performed on several types of air cassettes. Light microscopy at a 400 to 1000x magnification is used for air cassette sample analysis. SanAir always analyzes 100% of the impacted slide.

Explanation of Background Densities

The background density of an air cassette aids in the overall interpretation of results as it indicates the level of background debris present (e.g. dander, pollen, fibers, insect parts, soot, fly ash, etc.). Excessive background debris may mask the presence of fungal spores thereby reducing the accuracy of the count. It may also serve as an alert that the volume of air pulled was too high or too low. The following table explains background densities.

Air Cassette Density	Amount of Particulate on Slide	Explanation
1	Insignificant	Should not skew any counts
1+	Low	Should not skew any counts
2	Low to Moderate	Should not skew any counts
2+	Moderate to High	May cause occlusion of small spores
3	High	May cause occlusion of small to medium spores
3+	Very High	Will cause occlusion of spores
4	Overloaded	Level of particulate too high to perform analysis

A Note About the Fungal Spores

In some instances certain groups of fungi cannot be identified due to a lack of distinguishing characteristics. These fungi will be categorized as non-specified spores on the final report.

The genera *Aspergillus* and *Penicillium* are typically composed of small, round spores that are difficult to distinguish from each other; therefore, they are grouped into the category *Aspergillus / Penicillium*. Other fungi that produce spores of similar characteristics may also be placed into this category, including *Paecilomyces*, *Talaromyces*, and *Trichoderma*, among others.

Stachybotrys and *Memnoniella* spores are coated with a sticky lime-layer that may inhibit aerosolization.

Any genus of fungi detected on an air cassette with a high raw count (i.e. exceeding 500 spores) may be estimated. Any estimate higher than 12,000 spores will be reported as >12,000.

Understanding the Air Cassette Report

Each sample has 3 columns of information provided. The left is the raw count which is the number of spores for that fungal type detected on the trace. The middle column is the count per cubic meter (Count/m³) which is the raw count converted based on the total volume pulled for that sample. It represents the number of spores that should be expected in a cubic meter of air from the location in question *if* the spores were distributed evenly throughout the air. This column is helpful for interpreting results when the samples were pulled at different total volumes. In other words, the raw count of a cassette pulled at 75 liters should not be compared to the raw count of a cassette pulled at 150 liters because there may be higher counts associated with the higher volume. By comparing the Count/m³ columns the difference in volumes are accounted for.

The analytical sensitivity is the lowest spore count detectable with reasonable certainty, and it is calculated this way using a raw count of one. Keep in mind there are 1,000 liters in a cubic meter.

$$1 \times (1,000 / \text{Total Volume in Liters})$$

How to calculate the count per cubic meter:

$$\text{Raw Count} \times (1,000 / \text{Total Volume in Liters})$$

The last column on the right shows the percentage for which each spore type comprised the total spore count.

Understanding the Air Cassette Graph (If included in the final report)

The graph is a visual representation of the baseline sample (usually the outdoor air sample) compared individually against each indoor sample. Each spore type found on the indoor sample is compared to what was found outdoors per cubic meter.

The graph shows the percentile representation of each indoor spore count derived by dividing the indoor Count/m³ by the outdoor Count/m³. If the percentage is below 50% of the outside count, then the bar is below 50 on the chart, which corresponds to %Within 50% of Baseline Count/m³.+ If the percentage is between 50 and 100%, then the bar on the chart will stop between 50 and 100, which corresponds to %Count/m³ comparable to Baseline.+ If the percentage is greater than 100%, then the bar will be above 100 on the chart, which corresponds to %Count/m³ higher than Baseline.+

Each organism is given a threshold level for the Count/m³. If this threshold level is not met in an inside sample, then the organism will not be graphed on the chart. This is used to prevent the graph from showing every spore type that is commonly found outside and doesn't typically indicate a possible moisture problem inside. For example, most common outdoor spores (e.g. ascospores, basidiospores, and *Cladosporium*) have a threshold level of 100. Therefore, in order to show up on the chart, the inside Count/m³ must be above 100. On the other hand, fungi that may indicate water damage (e.g. *Stachybotrys*, *Ulocladium*, *Chaetomium*, *Memnoniella*, etc.) are given lower threshold levels. These fungi have a higher water activity value and therefore require more moisture to grow. *Stachybotrys* and *Chaetomium* have threshold values of 14 and 30, respectively, as even a low count of those types of spores may indicate an issue with excess moisture.

Keep in mind that this graph is to be used only as a tool in the inspection of a building. Visual examination and knowledge of water damage, past remediation, and weather conditions, among other elements, is essential in the decision regarding the indoor air quality of a building.

Assistance with Remediation Projects

more information pertaining to interpretation of results is available on our website www.sanair.com

For assistance in a remediation project you may consult the Institute of Inspection, Cleaning and Restoration Certification (IICRC) S500 and S520 protocols. The S500 is a reference guide for water-damage restoration and the S520 pertains specifically to mold remediation. Other standards and guidelines regarding Indoor Air Quality that may assist in remediation projects:

- AIHA (Recognition, Evaluation, and Control of Indoor Mold)
- AIHA (The Facts About Mold)
- NADCA (ACR 2006)
- IESO (Standards of Practice for the Assessment of Indoor Air Quality)
- EPA (Mold Remediation in Schools and Commercial Buildings)
- New York City Department of Health and Mental Hygiene (Guidelines on Assessment and Remediation of Fungi in Indoor Environments)

Disclaimer

SanAir Technologies Laboratory does not make contamination corrections to reports based upon analysis of laboratory and/or field blanks.

This report is the sole property of the client named on the SanAir Technologies Laboratory chain-of-custody. Results in the report are confidential information intended only for the use by the customer listed on the chain of custody (COC). Neither results nor reports will be discussed with or released to any third party without our clients' written permission. Final reports cannot be reproduced, except in full, without written authorization from SanAir. This report and any information contained within shall not be edited, altered, or modified in any way by any persons or agencies receiving, viewing, distributing, or otherwise possessing a copy of this final report. The laboratory reserves the right to perform amendments to any finalized report, of which shall supersede and make obsolete any previous editions. Such changes, modifications, additions, or deletions shall be effective immediately upon notice thereof, which may be given by means including but not limited to posting on the SanAir client portal website, electronic or conventional mail, or by any other means. The information provided in this report applies only to the samples submitted and is relevant only for the date, time, and location of sampling. The accuracy of the results of the analysis is dependent upon the method of sample procurement and information provided by the client on the COC. SanAir assumes no responsibility for the method of sample procurement. SanAir assumes no responsibility for information provided by the client on the COC such as project number, project name, collection dates, po number, special instructions, samples collected by technician name, sample numbers, sample identifications, sample type, selected analysis type, flow rate, total volume or area, and start stop times that may affect the validity of the results in this report. Evaluation reports are based solely on the sample(s) in the condition in which they arrived at the laboratory and on the information provided by the client on the COC. Sample(s) were received in good condition unless otherwise noted on the report. SanAir assumes no responsibility or liability for the manner in which the results are used or interpreted. SanAir will not provide any opinion on the safety of a building as visual inspection and knowledge of water damage, past remediation and weather conditions during sampling, among other elements, is essential in this decision. All samples are disposed of after 90 days unless otherwise requested by the client. SanAir is accredited by AIHA LAP, LLC in the EMLAP program. Refer to our accreditation certificate and scope or www.aihaaccreditedlabs.org for an up to date list of the Fields of Testing for which we are accredited.

This report does not constitute nor shall be used by the client to claim product, process, system, or person certification, approval, or endorsement by AIHA LAP, LLC, NVLAP, NELAC, NIST and/or any other U.S. governmental agencies; and may not be accredited by every local, state and federal regulatory agency.

LELAP Lab ID#05088

AIHA LAP, LLC Lab ID: LAP-162952



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Microbiology
Chain of Custody
 Form 68, Revision 7, 5/18/18

SanAir ID Number
 24064184

Company: FLI Environmental, Inc.	Project Number: 24-2539	Phone #: 781 251-0040
Address: 69 Bridge Street	Project Name: 90 CONCORD ST Ashland, MA	Phone #:
City, State, Zip: Dedham, MA 02026	Date Collected: 11-6-94	Fax #:
Samples Collected By: J Pike	P.O. Number:	Email: sclaus@flienv.com
Account #: 2799		Email: dmac@flienv.com

Sample Types		Analysis Types	Turn Around Time
AC	Air Cassette	A1 - Identification and Enumeration of Fungal spores, plus total dander, fiber, and pollen count	3/6/24/48 Hour
		A2 - Identification and Enumeration of Fungal spores only	3/6/24/48 Hour
T B S	Tape Bulk Swab	D1 - Direct Identification of Fungi	3/6/24/48 Hour
		D2 - Direct Identification of Mites, Insects, Pollen, etc.	3/6/24/48 Hour
		D3 - Direct Identification and Enumeration of Fungi	3/6/24/48 Hour
AP B S	Air Plate Bulk Swab	C1 - Culture Identification and Enumeration of Fungi only	5-10 Days
		C2 - Culture Identification and Enumeration of Bacteria only	2-4 Days
		C3 - Culture Identification and Enumeration of Fungi and Bacteria	5-10 Days
		C4 - Culture Identification and Enumeration of Thermophilic Bacteria with C2 or C3 analysis	2-4 or 5-10 Days
D	Dust	DA1 - Dust Mite Allergen Test	3/6/24/48 Hour

SanAir offers *Legionella* testing and other specialized culture analyses. Please call for details, COC and pricing.

Sample #	Sample Identification	Sample Type	Analysis Type(s)	Turn Around Time	Flow Rate (Liters/min)	Total Volume (L) or Area (in ²)	Time Start - Stop
1	Office Rm 184	AC	A2	48 HR	15L	75L	
2	Classroom Rm 167	AC	A2	48 HR	15L	75L	
3	Classroom Rm 145	AC	A2	48 HR	15L	75L	
4	Classroom Rm 154	AC	A2	48 HR	15L	75L	
5	Classroom Rm 149	AC	A2	48 HR	15L	75L	
6	Ambient	AC	A2	48 HR	15L	75L	

Special Instructions

Relinquished by	Date	Time	Received by	Date	Time
JPK	11-6-24		EDR	11/07/24	10:50 a.m.

If no technician is provided, then the primary contact for your account will be selected. Unless scheduled, the turnaround time for all samples received after 3 pm EST will be logged in the next business day. Weekend or holiday work must be scheduled ahead of time and is charged at 150% of the 3hr TAT or a minimum charge of \$150. A courier charge will be applied for same day and one-day turnaround times for offsite work. SanAir covers Standard Overnight FedEx shipping. Shipments billed to SanAir with a faster shipping rate will result in additional charges.